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DICTIONARY FILE UPDATES: 10 FEB 2002 HIGHEST RN 391197-12-9

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TSCA INFORMATION NOW CURRENT THROUGH July 7, 2001

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Crossover limits have been increased. See HELP CROSSOVER for details.

Calculated physical property data is now available. See HELP PROPERTIES
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Registry File, for complete details:
<http://www.cas.org/ONLINE/STN/STNOTES/stnotes27.pdf>

The P indicator for Preparations was not generated for all of the
CAS Registry Numbers that were added to the H/Z/CA/CAplus files between
12/27/01 and 1/23/02. Use of the P indicator in online and SDI searches
during this period, either directly appended to a CAS Registry Number
or by qualifying an L-number with /P, may have yielded incomplete results.
As of 1/23/02, the situation has been resolved. Also, note that searches
conducted using the PREP role indicator were not affected.

Customers running searches and/or SDIs in the H/Z/CA/CAplus files
incorporating CAS Registry Numbers with the P indicator between 12/27/01
and 1/23/02, are encouraged to re-run these strategies. Contact the
CAS Help Desk at 1-800-848-6533 in North America or 1-614-447-3698,
worldwide, or send an e-mail to help@cas.org for further assistance or to
receive a credit for any duplicate searches.

=> d ide can 11

L1 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2002 ACS
RN 7439-95-4 REGISTRY
CN Magnesium (8CI, 9CI) (CA INDEX NAME)
OTHER NAMES:
CN JIS 1
CN Magnesium element
CN PK 31
CN PK 31 (magnesium)
CN Rieke's active magnesium
DR 14147-08-1, 67208-78-0, 199281-20-4, 298688-48-9
MF Mg
CI COM
LC STN Files: ANABSTR, BIOBUSINESS, BIOSIS, BIOTECHNO, CA, CABA, CANCERLIT,
CAPLUS, CASREACT, CBNB, CEN, CHEMCATS, CHEMINFORMRX, CHEMLIST, CIN,
CSCHEM, CSNB, DDFU, DEtherm*, DRUGU, EMBASE, ENCOMPLIT, ENCOMPLIT2,
ENCOMPPAT, ENCOMPPAT2, HSDB*, IFICDB, IFIPAT, IFIUDB, IPA, MEDLINE,
MRCK*, MSDS-OHS, NAPRALERT, NIOSHTIC, TOXCENTER, TOXLIT, ULIDAT, USPAT2,
USPATFULL, VETU, VTB
(*File contains numerically searchable property data)
Other Sources: DSL**, EINECS**, TSCA**
(**Enter CHEMLIST File for up-to-date regulatory information)

Mg

149428 REFERENCES IN FILE CAPLUS (1967 TO DATE)

REFERENCE 1: 136:111823
REFERENCE 2: 136:111815
REFERENCE 3: 136:111798
REFERENCE 4: 136:111791
REFERENCE 5: 136:111768
REFERENCE 6: 136:111766
REFERENCE 7: 136:111759
REFERENCE 8: 136:111263
REFERENCE 9: 136:111062
REFERENCE 10: 136:111025

=> d ide can 12

L2 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2002 ACS
RN 7786-30-3 REGISTRY
CN Magnesium chloride (MgCl₂) (9CI) (CA INDEX NAME)
OTHER CA INDEX NAMES:
CN **Magnesium chloride (6CI, 7CI, 8CI)**
OTHER NAMES:
CN Aerotex Accelerator MX
CN Catalyst G
CN Magnesium dichloride
CN Magnogene
CN TMT 2
DR 12285-34-6, 77069-22-8
MF Cl₂ Mg
CI COM
LC STN Files: ADISNEWS, AGRICOLA, ANABSTR, BIOBUSINESS, BIOSIS, BIOTECHNO,
CA, CANCERLIT, CAOLD, CAPLUS, CASREACT, CBNB, CEN, CHEMCATS,
CHEMINFORMRX, CHEMLIST, CIN, CSCHEM, CSNB, DDFU, DETHERM*, DIOGENES,
DRUGU, EMBASE, ENCOMPLIT, ENCOMPLIT2, ENCOMPPAT, ENCOMPPAT2, GMELIN*,
HSDB*, IFICDB, IFIPAT, IFIUDB, MEDLINE, MRCK*, MSDS-OHS, NIOSHTIC,
PDLCOM*, PHAR, PIRA, PROMT, RTECS*, TOXCENTER, TOXLIT, TULSA, USAN,
USPAT2, USPATFULL, VETU, VTB
(*File contains numerically searchable property data)
Other Sources: DSL**, EINECS**, TSCA**
(**Enter CHEMLIST File for up-to-date regulatory information)

Cl-Mg-Cl

20861 REFERENCES IN FILE CA (1967 TO DATE)
·504 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA
20880 REFERENCES IN FILE CAPLUS (1967 TO DATE)
13 REFERENCES IN FILE CAOLD (PRIOR TO 1967)

REFERENCE 1: 136:111828
REFERENCE 2: 136:111556
REFERENCE 3: 136:109302
REFERENCE 4: 136:109078

REFERENCE 5: 136:107514
 REFERENCE 6: 136:107453
 REFERENCE 7: 136:106236
 REFERENCE 8: 136:105413
 REFERENCE 9: 136:104876
 REFERENCE 10: 136:104608

=> d his

(FILE 'HOME' ENTERED AT 16:22:44 ON 11 FEB 2002)
 SET COST OFF

FILE 'REGISTRY' ENTERED AT 16:22:56 ON 11 FEB 2002
 E MAGNESIUM/CN

L1 1 S E3
 E MAGNESIUM CHLORIDE/CN
 L2 1 S E3

FILE 'HCAPLUS' ENTERED AT 16:23:32 ON 11 FEB 2002
 E ACTIN/CT

E E3+ALL
 L3 1 S E1
 E E2+ALL
 L4 19413 S E2
 L5 1133 S E2 (L) G
 L6 1134 S L3,L5
 L7 4805 S ACTIN (L) G
 L8 4805 S L6,L7
 L9 36371 S ACTIN
 L10 168657 S L1 OR L2
 L11 47463 S MAGNESIUM CHLORIDE OR MGCL2 OR MAGNESIU
 L12 895 S L3-L9 AND L10,L11
 L13 36 S L12 AND ?CRYST?
 L14 130 S L3-L9 AND (PARACRYST? OR PARA(L)?CRYST?)
 L15 24 S L14 AND L10,L11
 L16 36 S L13,L15
 L17 125 S ACTINS/CW (L) PREP/RL
 L18 1 S L16 AND L17
 E HARTMAN J/AU
 L19 22 S E3,E11
 E HARTMAN JAMES/AU
 L20 9 S E3,E8,E9
 E MALIK F/AU
 L21 193 S E3-E12
 E SAKOZIC R/AU
 E SAKOWIC R/AU
 L22 23 S E10,E12
 E FINER J/AU
 L23 .16 S E3,E6,E9,E10
 L24 7 S L3-L9,L17 AND L19-L23
 L25 13 S L16 AND (FORMATION OR ISOLATION OR CHARACTERIZATION OR POLYMO
 L26 12 S L25 NOT ASCARIS/TI
 L27 13 S L18,L26
 L28 12 S L27 AND (MAGNESIUM OR MGCL2 OR MG###)
 L29 19 S L24,L28 AND L1-L28
 L30 1 S L29 AND L17
 L31 18 S L29 NOT L30

FILE 'REGISTRY' ENTERED AT 16:44:38 ON 11 FEB 2002

=> fil hcplus

FILE 'HCAPLUS' ENTERED AT 16:44:50 ON 11 FEB 2002
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FILE COVERS 1907 - 8 Feb 2002 VOL 136 ISS 7
FILE LAST UPDATED: 30 Jan 2002 (20020130/ED)

This file contains CAS Registry Numbers for easy and accurate substance identification.

CAS roles have been modified effective December 16, 2001. Please check your SDI profiles to see if they need to be revised. For information on CAS roles, enter HELP ROLES at an arrow prompt or use the CAS Roles thesaurus (/RL field) in this file.

The P indicator for Preparations was not generated for all of the CAS Registry Numbers that were added to the CAS files between 12/27/01 and 1/23/02. As of 1/23/02, the situation has been resolved. Searches and/or SDIs in the H/Z/CA/CAplus files incorporating CAS Registry Numbers with the P indicator executed between 12/27/01 and 1/23/02 may be incomplete. See the NEWS message on this topic for more information.

=> d 131 all hitstr tot

L31 ANSWER 1 OF 18 HCAPLUS COPYRIGHT 2002 ACS
AN 1999:5376 HCAPLUS
DN 130:220029
TI Use of optical traps in single-molecule study of nonprocessive biological motors
AU Mehta, A. D.; Finer, J. T.; Spudich, J. A.
CS Department of Biochemistry, Stanford University School of Medicine,
Stanford, CA, 94305, USA
SO Methods Enzymol. (1998), 298(Molecular Motors and the Cytoskeleton, Part
B), 436-459
CODEN: MENZAU; ISSN: 0076-6879
PB Academic Press
DT Journal
LA English
CC 9-5 (Biochemical Methods)
AB The authors describe the single-mol. measurements, using the gliding assay as the point of departure. The authors first discussed prepn. of proteins, coverslips, and labeled polystyrene beads for use in optical trapping. Then they provide a sketch of instrument design. Finally, they focus on exptl. conditions and data anal. The problems in identifying single-mol. binding events and methods developed to overcome them are also reviewed. (c) 1998 Academic Press.
ST optical trap single mol study biol motor
IT Filaments
Muscle
Optical traps
(use of optical traps in single-mol. study of nonprocessive biol.
motors)

IT Actins

Proteins (general), biological studies

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(use of optical traps in single-mol. study of nonprocessive biol.
motors)

RE.CNT 29 THERE ARE 29 CITED REFERENCES AVAILABLE FOR THIS RECORD

- RE
- (1) Dupuis, D; J Muscle Res Cell Motil 1997, V18, P17 MEDLINE
 - (2) Finer, J; Biophys J 1995, V68, P291s HCAPLUS
 - (3) Finer, J; Nature 1994, V368, P113 HCAPLUS
 - (4) Guilford, W; Biophys J 1997, V72, P1006 HCAPLUS
 - (5) Harada, Y; Cell Motil Cytoskelet 1988, V10, P71 HCAPLUS
 - (6) Harada, Y; J Molec Biol 1990, V216, P49 HCAPLUS
 - (7) Hua, W; Nature 1997, V388, P390 HCAPLUS
 - (8) Ishijima, A; Biochem Biophys Res Commun 1994, V199, P1057 HCAPLUS
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 - (10) Kellermayer, M; Science 1997, V276, P1112 HCAPLUS
 - (11) Kron, S; Methods Enzymol 1991, V196, P399 HCAPLUS
 - (12) Kron, S; Proc Natl Acad Sci USA 1986, V83, P6272 HCAPLUS
 - (13) Mehta, A; Methods Cell Biol 1998, V55, P47 MEDLINE
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 - (15) Mehta, A; Proc Natl Acad Sci USA 1997, V94, P7927 HCAPLUS
 - (16) Molloy, J; Biophys J 1995, V68, P298s HCAPLUS
 - (17) Molloy, J; Biophys J 1997, V72, P984 HCAPLUS
 - (18) Molloy, J; Nature 1995, V378, P209 HCAPLUS
 - (19) Nishizaka, T; Nature 1995, V377, P251 HCAPLUS
 - (20) Reif, M; Science 1997, V276, P1109
 - (21) Schnitzer, M; Nature 1997, V388, P386 HCAPLUS
 - (22) Simmons, R; Biophys J 1996, V70, P1813 HCAPLUS
 - (23) Suzuki, N; Biophys J 1996, V70, P401 HCAPLUS
 - (24) Svoboda, K; Nature 1993, V365, P721 HCAPLUS
 - (25) Svoboda, K; PNAS 1994, V91, P11782 HCAPLUS
 - (26) Tskhovrebova, L; Nature 1997, V387, P308 HCAPLUS
 - (27) Visscher, K; IEEE J Select Topics Quantum Electron 1996, V2, P1066 HCAPLUS
 - (28) Yanagida, T; Nature 1985, V316, P366 HCAPLUS
 - (29) Yin, H; Science 1995, V270, P1653 HCAPLUS

L31 ANSWER 2 OF 18 HCAPLUS COPYRIGHT 2002 ACS

AN 1998:288612 HCAPLUS

DN 129:91799

TI A six-module human nebulin fragment bundles actin
filaments and induces actin polymerization

AU Gonsior, Sabine M.; Gautel, Mathias; Hinssen, Horst

CS Biochemical Cell Biology Group, University of Bielefeld, Bielefeld, 33615,
Germany

SO J. Muscle Res. Cell Motil. (1998), 19(3), 225-235

CODEN: JMRMD3; ISSN: 0142-4319

PB Chapman & Hall

DT Journal

LA English

CC 6-1 (General Biochemistry)

AB The authors have investigated the interaction of a 6-repeat recombinant

human nebulin fragment (S6R2R7) with F-actin, with Mg2

+-induced actin paracrystals, and G-

actin, resp. This fragment corresponds to super-repeat 6, repeat

2 to 7 of human nebulin, and is located in the N-terminal part of the
super-repeat region of the nebulin mol. The S6R2R7 fragment included an
immuno-tag of three amino-acid residues (EEF) at one end which was
detectable by a monoclonal anti-tubulin YL1/2. By a cosedimentation
assay, interaction between F-actin and S6R2R7 was obsd.Electron microscopy revealed the formation of large bundle-like aggregates
contg. highly parallelized actin filaments, apparently caused by
actin bundling of the nebulin fragment. Compared with Mg2
+-induced actin paracrystals where the helixes of the
actin filaments are arranged in register, the filaments in the
actin-nebulin bundles seem to be packed in a different way and

show no obvious periodicity. The bundles were also visible in the light microscope, and immunofluorescence microscopy revealed binding of the nebulin fragment S6R2R7 to both preformed Mg²⁺ paracrystals and to F-actin. The authors also analyzed the effect of S6R2R7 on actin under non-polymg. conditions by cosedimentation assays and pyrene actin fluorimetry, as well as fluorescence microscopy and electron microscopy. Nebulin-induced actin polymn. was obsd. with an enhancement of the nucleation step indicating a stabilization of actin nuclei by S6R2R7. Light and electron microscopy revealed bundle-like actin-nebulin aggregates similar to those formed by pre-assembled F-actin and S6R2R7. Thus, even in the absence of salt, S6R2R7 promotes actin polymn. and induces formation of tightly packed actin filament bundles. It was assumed that the actin filaments are crosslinked by the nebulin fragments, indicating a rather low cooperativity of binding to a single filament.

ST nebulin fragment actin filament bundling; actin polymn
nebulin fragment

IT Quasicrystals

(Mg²⁺-induced actin paracrystals;
six-module human nebulin fragment bundles actin filaments and
induces actin polymn.)

IT Actin filament

Polymerization

(six-module human nebulin fragment bundles actin filaments
and induces actin polymn.)

IT F-actins

G-actins

Nebulin

RL: BPR (Biological process); BIOL (Biological study); PROC (Process)
(six-module human nebulin fragment bundles actin filaments
and induces actin polymn.)

IT 7439-95-4, Magnesium, biological studies

RL: BAC (Biological activity or effector, except adverse); BIOL
(Biological study)

(Mg²⁺-induced actin paracrystals;
six-module human nebulin fragment bundles actin filaments and
induces actin polymn.)

IT 7439-95-4, Magnesium, biological studies

RL: BAC (Biological activity or effector, except adverse); BIOL
(Biological study)

(Mg²⁺-induced actin paracrystals;
six-module human nebulin fragment bundles actin filaments and
induces actin polymn.)

RN 7439-95-4 HCPLUS

CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 3 OF 18 HCPLUS COPYRIGHT 2002 ACS

AN 1997:479465 HCPLUS

DN 127:187682

TI Detection of single-molecule interactions using correlated thermal diffusion

AU Mehta, A. D.; Finer, J. T.; Spudich, J. A.

CS Departments Biochemistry Developmental Biology, Beckman Center, Stanford University School Medicine, Stanford, CA, 94305, USA

SO Proc. Natl. Acad. Sci. U. S. A. (1997), 94(15), 7927-7931

CODEN: PNASA6; ISSN: 0027-8424

PB National Academy of Sciences

DT Journal

LA English

CC 9-5 (Biochemical Methods)

AB Observation of discrete, single-mol. binding events allows one to bypass assumptions required to infer single-mol. properties from studies of ensembles of mols. Optically trapped beads and glass microneedles have been applied to detect single-mol. binding events, but it remains difficult to identify signs of binding events given the large displacements induced by thermal forces. Here, we exploit thermal diffusion by using correlation between motion of optically trapped beads attached to both ends of a single **actin** filament to track binding events of individual myosin mols. We use correlated diffusion to measure the stiffness of a single myosin mol. and est. its thermal fluctuation in a poststroke state as comparable in amplitude to the measured stroke distance. The use of correlated diffusion to measure kinetics of single-mol. interactions and the stiffness of the interacting moieties should be applicable to any pair of interacting mols., and not limited to biol. motors.

ST thermal diffusion binding mol detection; myosin binding **actin**
bead movement detection

IT **Actins**

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(beads attached to; detection of myosin binding to single filament
actin by movement of micrometer sized beads)

IT Thermal diffusion

(detection of single-mol. interactions using correlated thermal diffusion)

IT **Myosins**

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(detection of single-mol. interactions using correlated thermal diffusion)

L31 ANSWER 4 OF 18 HCPLUS COPYRIGHT 2002 ACS

AN 1996:179980 HCPLUS

DN 124:223983

TI Single myosin molecule mechanics (muscle contraction, **actin** filament)

AU **Finer, Jeffrey Todd**

CS Stanford Univ., Stanford, CA, USA

SO (1996) 234 pp. Avail.: Univ. Microfilms Int., Order No. DA9602876

From: Diss. Abstr. Int., B 1996, 56(10), 5468

DT Dissertation

LA English

CC 6-1 (General Biochemistry)

AB Unavailable

ST myosin mechanics muscle contraction **actin** filament

IT Muscle

(contraction; single myosin mol. mechanics in relation to muscle contraction and **actin** filament)

IT **Myosins**

RL: PEP (Physical, engineering or chemical process); PROC (Process)
(single myosin mol. mechanics in relation to muscle contraction and **actin** filament)

IT **Actins**

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(F-, single myosin mol. mechanics in relation to muscle contraction and **actin** filament)

IT **Microfilament**

(thin filament, single myosin mol. mechanics in relation to muscle contraction and **actin** filament)

L31 ANSWER 5 OF 18 HCPLUS COPYRIGHT 2002 ACS

AN 1995:467785 HCPLUS

DN 122:259108

TI Characterization of single **actin-myosin** interactions

AU **Finer, Jeffrey T.; Mehta, Amit D.; Spudich, James A.**

CS Dep. Biochem. Dev. Biol., Stanford Univ. Med. Cent., Stanford, CA, 94305,
USA

SO Biophys. J. (1995), 68(4, Suppl.), 291s-7s

DT CODEN: BIOJAU; ISSN: 0006-3495
 LA Journal
 English
 CC 6-3 (General Biochemistry)
 AB The feedback-enhanced laser trap assay (Finer et al., 1994) allows the measurement of force and displacement produced by single myosin mols. interacting with an actin filament suspended in soln. by two laser traps. The av. displacement of 11 nm at low load and the av. force of 4 pN near isometric conditions are consistent with the conventional swinging cross-bridge model of muscle contraction (Huxley, 1969). The durations of single actin-myosin interactions at low load, 3-7 ms, suggest a relatively small duty ratio. Event durations can be increased either by reducing the ATP concn. until ATP binding is rate-limiting or by lowering the temp. For sufficiently long interactions near isometric conditions, low frequency force fluctuations were obsd. within the time frame of a single event. Single myosin events can be measured at ionic strengths that disrupt weak binding actomyosin interactions, supporting the postulate of distinct weak and strong binding states. Myosin-generated force and displacement were measured simultaneously against several different loads to generate a force-displacement curve. The linear appearance of this curve suggests that the myosin powerstroke is driven by the release of a strained linear elastic element with a stiffness of approx. 0.4 pN nm⁻¹.
 ST actin myosin interaction
 IT Molecular association
 (single actin-myosin interactions)
 IT Actins
 Myosins
 RL: BPR (Biological process); BIOL (Biological study); PROC (Process)
 (single actin-myosin interactions)
 IT 56-65-5, 5'-Atp, biological studies
 RL: BAC (Biological activity or effector, except adverse); BIOL
 (Biological study)
 (single actin-myosin interactions in presence of)

 L31 ANSWER 6 OF 18 HCPLUS COPYRIGHT 2002 ACS
 AN 1994:453307 HCPLUS
 DN 121:53307
 TI Single myosin molecule mechanics: piconewton forces and nanometer steps
 AU Finer, Jeffrey T.; Simmons, Robert M.; Spudich, James A.
 CS Sch. Med., Stanford Univ., Stanford, CA, 94305, USA
 SO Nature (London) (1994), 368(6467), 113-19
 CODEN: NATUAS; ISSN: 0028-0836
 DT Journal
 LA English
 CC 9-5 (Biochemical Methods)
 Section cross-reference(s): 13
 AB A new in vitro assay using a feedback enhanced laser trap system allows direct measurement of force and displacement that results from the interaction of a single myosin mol. with a single suspended actin filament. Discrete stepwise movements averaging 11 nm were seen under conditions of low load, and single force transients averaging 3-4 pN were measured under isometric conditions. The magnitudes of the single forces and displacements are consistent with predictions of the conventional swinging-crossbridge model of muscle contraction.
 ST myosin mechanics actin filament method; muscle contraction
 myosin mechanics actin method
 IT Muscle
 (contraction of, myosin single mol. movement and forces on
 actin filament anal. by, optical trap method in relation to)
 IT Actins
 RL: ANST (Analytical study)
 (filament, myosin single mol. movement and forces on, optical trap
 method for detn. of)
 IT Force
 (in myosin single mol. on actin filament, optical trap method

for anal. of)

IT Myosins
 RL: ANST (Analytical study)
 (mechanics of single mol. of, on actin filament, optical trap method for detn. of movement and forces in)

IT Microfilament
 (thin filament, actin, myosin single mol. movement and forces on, optical trap method for detn. of)

L31 ANSWER 7 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1994:100841 HCAPLUS
 DN 120:100841
 TI In vitro methods for measuring force and velocity of the actin-myosin interaction using purified proteins
 AU Warrick, Hans M.; Simmons, Robert M.; Finer, Jeffrey T.; Uyeda, Taro Q. P.; Chu, Steven; Spudich, James A.
 CS Sch. Med., Stanford Univ., Stanford, CA, 94305, USA
 SO Methods Cell Biol. (1993), 39(Motility Assays for Motor Proteins), 1-21
 CODEN: MCBLAG; ISSN: 0091-679X
 DT Journal; General Review
 LA English
 CC 9-0 (Biochemical Methods)
 Section cross-reference(s): 6, 13
 AB A review with many refs. Prepn. of in vitro motility assay components, in vitro assay for myosin velocity and for myosin force in the motility assay, components of the optical trap system, and future directions related to the title method are included.
 ST actin myosin interaction review
 IT Myosins
 RL: ANST (Analytical study)
 (interactions of, with actins, force and velocity measurement of)
 IT Actins
 RL: ANST (Analytical study)
 (interactions of, with myosins, force and velocity measurement of)

L31 ANSWER 8 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1991:138654 HCAPLUS
 DN 114:138654
 TI Nucleotide specificity of the enzymic and motile activities of dynein, kinesin, and heavy meromyosin
 AU Shimizu, Takashi; Furusawa, Kiyotaka; Ohashi, Shinichi; Toyoshima, Yoko Y.; Okuno, Makoto; Malik, Fady; Vale, Ronald D.
 CS Res. Inst. Polym. Text., Tsukuba, 305, Japan
 SO J. Cell Biol. (1991), 112(6), 1189-97
 CODEN: JCLBA3; ISSN: 0021-9525
 DT Journal
 LA English
 CC 7-3 (Enzymes)
 Section cross-reference(s): 6
 AB The substrate specificities of dynein, kinesin, and myosin substrate turnover activity and cytoskeletal filament-driven translocation were examd. using 15 ATP analogs. The dyneins were more selective in their substrate utilization than bovine brain kinesin or muscle heavy meromyosin, and even different types of dyneins, such as 14 S and 22 S dynein from Tetrahymena cilia and the .beta.-heavy chain-contg. particle from the outer-arm dynein of sea urchin flagella, could be distinguished by their substrate specificities. Although bovine brain kinesin and muscle heavy meromyosin both exhibited broad substrate specificities, kinesin-induced microtubule translocation varied over a 50-fold range in speed among the various substrates, whereas heavy meromyosin-induced actin translocation varied only by 4-fold. With both kinesin and heavy meromyosin, the relative velocities of filament translocation did not correlate well with the relative filament-activated substrate turnover rates. Furthermore, some ATP analogs that did not support the filament translocation exhibited filament-activated substrate turnover rates.

Filament-activated substrate turnover and power prodn., therefore, appear to become uncoupled with certain substrates. In conclusion, the substrate specificities and coupling to motility are distinct for different types of mol. motor proteins. Such nucleotide fingerprints of enzymic activities of motor proteins may prove useful as a tool for identifying what type of motor is involved in powering a motility-related event that can be reconstituted in vitro.

ST cytoskeleton filament motility ATPase ATP analog; dynein motility ATPase ATP analog; kinesin motility ATPase ATP analog; meromyosin motility ATPase ATP analog

IT Cilia
(motility of, ATP analogs specificity of, ATPase specificity in relation to)

IT Michaelis constant
(of ATPase, of microtubule-dynein system)

IT Microtubule
(translocation of, in system with dyneins, ATP analog specificity of, ATPase specificity in relation to)

IT Dyneins
RL: BIOL (Biological study)
(14 S, ATPase and motile activity of, ATP analog specificity of)

IT Dyneins
RL: BIOL (Biological study)
(22 S, ATPase and motile activity of, ATP analog specificity of)

IT Meromyosins
RL: BIOL (Biological study)
(heavy, ATPase of, ATP analogs specificity of)

IT Meromyosins
RL: BIOL (Biological study)
(heavy, acto-, ATPase and motile activity of, ATP analog specificity of)

IT Proteins, specific or class
RL: BIOL (Biological study)
(kinesins, ATPase and motile activity of, ATP analog specificity of)

IT Biological transport
(translocation, filament-driven, in cytoskeleton, ATP analog specificity of, dyneins and kinesin and heavy meromyosin ATPase specificity in relation to)

IT 56-65-5, 5'-ATP, biological studies 73-04-1, 3'-Deoxy ATP 1927-31-7,
2'-Deoxy ATP 2677-93-2 3130-39-0 16409-13-5, Formycin
5'-triphosphate 23197-96-8 23567-97-7 24027-80-3 35094-46-3,
Adenosine 5'-O-(3-thiotriphosphate) 37482-17-0 53696-59-6, 8-Azido ATP
58976-48-0 58976-49-1 59261-35-7 59261-36-8
RL: BIOL (Biological study)
(ATPase and motile activities of dynein and kinesin and meromyosin specificity for)

IT 9000-83-3, ATPase
RL: BIOL (Biological study)
(of dyneins and kinesins and meromyosin, ATP analogs specificity of, filament motility in relation to)

L31 ANSWER 9 OF 18 HCPLUS COPYRIGHT 2002 ACS
AN 1990:567651 HCPLUS
DN 113:167651
TI A polymorphism peculiar to bipolar actin bundles
AU Francis, Noreen R.; DeRosier, David J.
CS Rosenstiel Basic Med Sci. Res. Cent., Brandeis Univ., Waltham, MA, 02254,
USA
SO Biophys. J. (1990), 58(3), 771-6
CODEN: BIOJAU; ISSN: 0006-3495
DT Journal
LA English
CC 6-3 (General Biochemistry)
AB Both muscle and nonmuscle actins produced Mg paracrystals which were indistinguishable from one another.

Contrary to some previous reports, Ca^{2+} caused no change in filament organization for either type of actin. The most ordered paracrystals consisted of hexagonally packed filaments with opposite polarities. It is suggested that this mode of packing permits a form of disorder not previously described, which may account for some puzzling aspects of earlier observations and may prove useful in analyzing actin bundles formed, for example, with erythrocyte band 4.9 protein.

ST bipolar actin bundle polymorphism; magnesium
actin paracrystal polymorphism
IT Quasicrystals
(of actin and magnesium, polymorphism of)
IT Actins
RL: BIOL (Biological study)
(F-, paracrystals of, polymorphism of, magnesium in
relation to)
IT Organelle
(actin bundle, bipolar, organization of, actin
magnesium paracrystal polymorphism in relation to)
IT 7439-95-4D, Magnesium, actin filament
complexes
RL: BIOL (Biological study)
(paracrystals of, polymorphism of)
IT 7439-95-4D, Magnesium, actin filament
complexes
RL: BIOL (Biological study)
(paracrystals of, polymorphism of)
RN 7439-95-4 HCPLUS
CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 10 OF 18 HCPLUS COPYRIGHT 2002 ACS
AN 1988:90001 HCPLUS
DN 108:90001
TI Isolation and characterization of actin from
cultured BHK cells
AU Koffer, Anna; Dickens, Michael J.
CS MRC Cell Biophys. Unit, London, WC2B 5RL, UK
SO J. Muscle Res. Cell Motil. (1987), 8(5), 397-406
CODEN: JMRMD3; ISSN: 0142-4319
DT Journal
LA English
CC 6-3 (General Biochemistry)
AB Cytoplasmic actin from cultured fibroblasts has been purified to
homogeneity and characterized with respect to its polymn. and structure.
It was qual. similar to muscle actin in all respects, but
significant quant. differences in its properties were demonstrated.
Although BHK actin did not polymerize in unfractionated
cytoplasmic exts., the purified BHK actin polymd. into filaments
both in the presence of Mg and Ca. The crit. concn., measured
by the DNase I inhibition assay and by fluorimetry, was the same as that
of muscle actin both in Mg and Ca. Polymn. of
pyrene-labeled BHK and muscle actin was followed by fluorimetry.
Significant differences in kinetics were found under both ionic conditions
tested. In the absence of Mg^{2+} (0.2 mM CaCl_2 , 85 mM KCl), BHK
actin polymd. at a much slower rate than did muscle actin
. In the presence of Mg and EGTA, the nucleation phase for BHK
actin polymn. was shorter than that for muscle actin and
the kinetics of polymn. was different. The structure of BHK actin
filaments in the electron micrographs was very similar to that of muscle
actin. In high concns. of Mg, BHK actin
formed paracrystals which had the same appearance as muscle

actin paracrystals. However, Ca-induced formation of actin paracrystals required higher concn. of Ca²⁺ for BHK actin than for muscle actin (12 mM and 8 mM, resp.). These results suggest differences in divalent cation binding to both high- and low-affinity sites of the two actins.

ST BHK cell actin; calcium binding actin cytoplasm;
magnesium binding actin cytoplasm; polymn actin

IT BHK cell
Cytoplasm
(actin of, of animal cell, isolation and polymn. and structure of, muscle actin comparison with)

IT **Actins**
RL: BIOL (Biological study)
(of BHK cell, isolation and polymn. and structure of, muscle actins comparison with)

IT Animal cell line
(BHK, actin of, isolation and polymn. and structure of, muscle actins comparison with)

IT **Actins**
RL: BIOL (Biological study)
(G-, of BHK cells, polymn. of, kinetics of)

IT 67-42-5, EGTA
RL: BIOL (Biological study)
(actin of BHK cell polymn. in magnesium presence acceleration by)

IT 7439-95-4, Magnesium, biological studies 7440-70-2,
Calcium, biological studies
RL: BIOL (Biological study)
(actin of BHK cell polymn. in presence of, kinetics of)

IT 7439-95-4, Magnesium, biological studies
RL: BIOL (Biological study)
(actin of BHK cell polymn. in presence of, kinetics of)

RN 7439-95-4 HCPLUS
CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 11 OF 18 HCPLUS COPYRIGHT 2002 ACS
 AN 1983:554040 HCPLUS
 DN 99:154040
 TI Structural studies of F-actin
 AU Egelman, Edward H.; DeRosier, David J.
 CS Biophys. Rosenst. Res. Cent., Brandeis Univ., Waltham, MA, USA
 SO Actin: Struct. Funct. Muscle Non-Muscle Cells, Proc. Int. Semin., Int. Congr. Biochem., 12th (1983), Meeting Date 1982, 17-24. Editor(s): Dos Remedios, Cristobal G.; Barden, Julian A. Publisher: Academic, North Ryde, Australia.
 CODEN: 50FOAW
 DT Conference
 LA English
 CC 6-3 (General Biochemistry)
 AB A model of the actin filament, developed from studies of isolated neg. stained F-actin, is quite consistent with images of neg. stained angle layered aggregates and freeze-etched single filaments. Further, the transform of the model agrees with obsd. x-ray patterns of muscle and of actin gels. All of these patterns show that the mass of the actin subunit is oriented approx. along the 59 .ANG. helix. Finally, by treating Mg²⁺ paracrystals as deformed angle layered aggregates, the obsd. appearance of paracrystals was simulated and a certain class of actin models were explained as arising from an artifact of superposition.
 ST actin magnesium paracrystal model

IT Microfilament and Microtubule
 (of actin, magnesium-induced, structure of, model
 for)
 IT **Actins**
 RL: BIOL (Biological study)
 (F-, magnesium-induced paracrystals of, structure
 of, model for)
 IT 7439-95-4, uses and miscellaneous
 RL: USES (Uses)
 (actin paracrystals induced by, structure of, model
 for)
 IT 7439-95-4, uses and miscellaneous
 RL: USES (Uses)
 (actin paracrystals induced by, structure of, model
 for)
 RN 7439-95-4 HCAPLUS
 CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 12 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1982:157741 HCAPLUS
 DN 96:157741
 TI Purification and characterization of tropomyosin from bovine thyroid
 AU Kobayashi, Ryoji; Tawata, Masato; Mace, Myles L., Jr.; Bradley, William A.; Field, James B.
 CS Diabetes Res. Cent., St. Luke's Episcopal Hosp., Houston, TX, 77030, USA
 SO Biochim. Biophys. Acta (1982), 702(2), 220-32
 CODEN: BBACAO; ISSN: 0006-3002.
 DT Journal
 LA English
 CC 6-3 (General Biochemistry)
 AB A tropomyosin was purified from bovine thyroid and its properties compared with those of rabbit skeletal muscle tropomyosin. Thyroid tropomyosin was sepd. from contaminating vascular smooth muscle tropomyosin by hydroxylapatite chromatog. Thyroid tropomyosin resembles tropomyosin from other nonmuscle cells in regard to subunit size, mobility on SDS-polyacrylamide gels in the presence and absence of 6M urea, amino acid compn., and morphol. Thyroid tropomyosin has a subunit mol. wt. of 30,000 and forms Mg²⁺ paracrystals with an axial period of 345 .ANG., whereas paracrystal periodicities of muscle tropomyosins are 400 .ANG.. The amino acid compn. of thyroid tropomyosin is very similar to that of other nonmuscle cell tropomyosins. However, thyroid tropomyosin differs from other nonmuscle cell tropomyosins in its ability to bind to actin and troponin. Both thyroid and muscle tropomyosins bind to actin in a similar ratio of 1 tropomyosin/6-7 actin monomers at satn. The binding of tropomyosin to F-actin is strongly dependent on the Mg²⁺ concn. With thyroid tropomyosin, binding begins at 1 mM and is complete at .apprx.4-5 mM Mg²⁺, whereas with muscle tropomyosin, binding is initiated at 1 mM Mg²⁺ and reaches satn. at 2-3 mM Mg²⁺. At satn., both thyroid and muscle tropomyosins bind to the same binding site(s) on actin filaments with similar affinity. In contrast to platelet tropomyosin, thyroid tropomyosin binds to skeletal muscle troponin and troponin T. One-dimensional peptide maps of thyroid and rabbit skeletal muscle tropomyosin are distinctly different from each other. The air oxidn. of thyroid tropomyosin yields covalently linked dimers similar to skeletal muscle tropomyosin dimers. In contrast to muscle tropomyosins, [32P]phosphate is not incorporated into thyroid tropomyosin.
 ST tropomyosin thyroid gland
 IT Tropomyosins

RL: BIOL (Biological study)
 (of thyroid, purifn. and properties of)
 IT Amino acids, biological studies
 RL: BIOL (Biological study)
 (of tropomyosin, of thyroid)
 IT Thyroid gland, composition
 (tropomyosin of)
 IT Troponins
 RL: BIOL (Biological study)
 (tropomyosin of thyroid binding to)
 IT **Actins**
 RL: BIOL (Biological study)
 (F-, tropomyosin of thyroid binding to)
 IT Troponins
 RL: BIOL (Biological study)
 (T, tropomyosin of thyroid binding to)
 IT **7439-95-4**, biological studies
 RL: BIOL (Biological study)
 (tropomyosin of thyroid binding to actin response to)
 IT **7439-95-4**, biological studies
 RL: BIOL (Biological study)
 (tropomyosin of thyroid binding to actin response to)
 RN 7439-95-4 HCAPLUS
 CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 13 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1981:402190 HCAPLUS
 DN 95:2190
 TI **Formation of actin paracrystals** from sea
 urchin egg extract under actin polymerizing conditions
 AU Mabuchi, Issei; Nonomura, Yoshiaki
 CS Coll. Gen, Educ., Univ. Tokyo, Tokyo, 153, Japan
 SO Biomed. Res. (1981), 2(2), 143-53
 CODEN: BRESD5
 DT Journal
 LA English
 CC 6-3 (General Biochemistry)
 AB A monomeric actin fraction was obtained from a high-speed
 supernatant of an ext. of unfertilized sea urchin (*Anthocidaris*
 crassipina) eggs by gel filtration chromatog. Ppts. formed on concn. of
 this fraction at 4.degree. were **paracrystals of actin**.
 These **paracrystals** contained **actin** and a
 56,000-mol.-wt. protein at a molar ratio of 4.8-5.0:1. The
 paracrystals dissolved in a low-ionic strength buffer soln. which
 depolymerizes **actin** and reformed on addn. of 0.1M KCl or 2 mM
 MgCl₂ at 0.degree.. Electron microscopy and optical diffraction
 studies showed that the **paracrystals** had transverse bands, the
 spacing of which was 1/3 of the distance between the crossover points of
 the 2 long-pitch right-handed helical strands of the **actin**
 filaments. Further, the **actin** filaments in the
 paracrystals had a helical configuration in which there were 41
 monomers/19 turns of the left-handed genetic **actin** helix. These
 structural properties may indicate that the **paracrystal** is an *in*
 vitro reconstituted microvillar **actin** core which is known to
 elongate on fertilization.
 ST **actin paracrystal** egg sea urchin
 IT *Anthocidaris crassispina*
 (actins of eggs of, structure of **paracrystals** of)
 IT Egg
 (actins of, **paracrystal** structure of, of sea
 urchin, microvillar core in relation to)

IT **Actins**
 RL: BIOL (Biological study)
 (paracrystals of, structure of, of sea urchin egg)

IT **Chains, chemical**
 (helical, of actin filaments in paracrystals)

L31 ANSWER 14 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1980:123710 HCAPLUS
 DN 92:123710
 TI Conformation changes of actin during formation of filaments and paracrystals and upon interaction with DNase I, cytochalasin B, and phalloidin
 AU Harwell, O. Daniel; Sweeney, Mary Lee; Kirkpatrick, Francis H.
 CS Sch. Med. Dent., Univ. Rochester, Rochester, NY, 14642, USA
 SO J. Biol. Chem. (1980), 255(3), 1210-20
 CODEN: JBCHA3; ISSN: 0021-9258
 DT Journal
 LA English
 CC 6-3 (General Biochemistry)
 AB Spin labels attached to rabbit muscle actin became more immobilized on conversion of actin from the G state to the F state with 50 mM KCl. Titrn. of G-actin with MgCl₂ produced F-actin-like EPR spectra between 2 and 5 mM, and F-actin filaments by electron microscopy. Higher concns. of MgCl₂ produced bundles of actin and eventually paracrystals, accompanied by further immobilization of spin labels. The effects of MgCl₂ and KCl were competitive: addn. of MgCl₂ to 50 mM converted F-actin (50 mM KCl) to paracryst. (P) actin; the reverse titrn. (0-200 mM KCl in the presence of 20 mM MgCl₂) was less complete. Addn. of DNase I to paracryst. actin gave the expected amorphous electron microg. pattern, and the actin was not sedimentable at 400,000 g (1 h). EPR showed that the actin was in the G conformation. Addn. of DNase I to paracryst. actin gave the F conformation (EPR) but the actin was G by electron microscopy. Phalloidin converted G-actin to F-actin, had no effect on F-actin, and converted P-actin to the F state by electron microscopy but maintained the P conformation by EPR. Cytochalasin B produced no effects observable by EPR or centrifugation but untwisted paracrystals into nets. Since actin retained its P conformation by EPR in 2 states which were morphol. not P, the P state is apparently a distinct conformation of the actin mol. and actin filaments aggregate to form bundles (and eventually paracrystals) when actin monomers can enter the P conformation.
 ST phalloidin actin conformation; cytochalasin B actin conformation; DNase I actin conformation; actin conformation filament paracrystal
 IT **Actins**
 RL: BIOL (Biological study)
 (conformational changes of, during G-to-F transition and modifier interaction)
 IT **Chains, chemical**
 (conformational transitions of, of actin during G-to-F transition and modifier interaction)
 IT 7447-40-7, properties 7786-30-3, properties 9003-98-9
 14930-96-2 17466-45-4
 RL: PRP (Properties)
 (conformation of actin response to)
 IT **7786-30-3, properties**
 RL: PRP (Properties)
 (conformation of actin response to)
 RN 7786-30-3 HCAPLUS
 CN Magnesium chloride (MgCl₂) (9CI) (CA INDEX NAME)

Cl-Mg-Cl

L31 ANSWER 15 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1980:106090 HCAPLUS
 DN 92:106090
 TI Depolymerization of actin in concentrated solutions of divalent metal chlorides
 AU Biro, E. N. A.; Ven'yaminov, S. Yu.
 CS Dep. Biochem., Eotvos Lorand Univ., Budapest, Hung.
 SO Acta Biochim. Biophys. Acad. Sci. Hung. (1979), 14(1-2), 31-42
 CODEN: ABBPAP; ISSN: 0001-5253
 DT Journal
 LA English
 CC 6-3 (General Biochemistry)
 AB Actin transferred to concd. (0.3-1.2M) MgCl₂ solns. depolymd. completely. When protected by a high excess of ATP, actin in this MgCl₂-depolymd. state was stable for several days in the cold. In the absence of excess ATP it slowly denatured. Chiroptical data and proteolysis expts. showed that MgCl₂-depolymd. actin is in a native, folded state, although its helix content is considerably decreased. By dissolving F-actin pellets or actin ptd. in the paracryst. state in concd. MgCl₂ solns. in the presence of ATP, very concd. (100-200 mg/mL) monomeric actin solns. were prep'd. CaCl₂ and MnCl₂ had similar effects, although these were not studied in detail.
 ST actin depolymn divalent metal chloride; magnesium chloride depolymn actin; calcium chloride depolymn actin; manganese chloride depolymn actin
 IT Chains, chemical (conformation of, of actin after depolymn. in concd. divalent metal chloride soln.)
 IT Actins
 RL: RCT (Reactant)
 (depolymn. of, in concd. divalent metal chloride soln., with ATP stabilization)
 IT Depolymerization
 (of actins, in concd. divalent metal chloride soln.)
 IT 56-65-5, biological studies
 RL: BIOL (Biological study)
 (actin depolymd. by concd. divalent metal chloride soln. stabilization by)
 IT 7773-01-5 7786-30-3, reactions 10043-52-4, reactions
 RL: BIOL (Biological study)
 (depolymn. of actin in concd. soln. of)
 IT 7786-30-3, reactions
 RL: BIOL (Biological study)
 (depolymn. of actin in concd. soln. of)
 RN 7786-30-3 HCAPLUS
 CN Magnesium chloride (MgCl₂) (9CI) (CA INDEX NAME)

Cl-Mg-Cl

L31 ANSWER 16 OF 18 HCAPLUS COPYRIGHT 2002 ACS
 AN 1978:524805 HCAPLUS
 DN 89:124805
 TI Interaction of actin with divalent cations.
 1. The effect of various cations on the physical state of actin
 AU Strzelecka-Golaszewska, Hanna; Prochniewicz, Ewa; Drabikowski, Witold
 CS Dep. Biochem. Nerv. Syst. Muscle, Nencki Inst. Exp. Biol., Warsaw, Pol.

SO Eur. J. Biochem. (1978), 88(1), 219-27
 CODEN: EJBCAI; ISSN: 0014-2956
 DT Journal
 LA English
 CC 6-3 (General Biochemistry)
 AB The effect of various divalent cations on the state of aggregation of actin monomers was studied at pH 7.6 by viscosity measurements, detn. of the protein sedimenting at high and low centrifugal forces, dephosphorylation of actin-bound ATP, and electron microscopy. The metal concn. dependence of the degree of actin polymn. in the presence of Ca²⁺, Mg²⁺, Sr²⁺, and Mn²⁺ was the same. All these cations produced typical double-stranded F-actin filaments. Ni²⁺ and Zn²⁺ induced polymn. at lower concns. than Mn²⁺ and alk. earth metals, but the resultant polymers had lower viscosities. Examn. in the electron microscope showed that Ni²⁺ produces typical F-actin filaments, which, however, tend to break into short fragments. In the presence of Zn²⁺ globular aggregates coexisting with the filaments were obsd. In the presence of Mn²⁺ or alk. earth metals at mM concns. the F-actin filaments assembled into netlike paracrystals which were transformed into side-by-side aggregates when the cation concn. was increased. The cation concn. dependences of polymn. and of paracrystal formation suggested that these 2 processes occur on binding of these cations to distinct classes of sites and that the order of affinities to sites of weaker binding, involved in the paracrystal formation, is as follows: Mn²⁺ > Ca²⁺ > Mg²⁺ = Sr²⁺. Unlike the other cations, Zn²⁺, at concns. higher than that necessary for max. polymn., caused pptn. of G-actin without formation of ordered structures.
 ST actin aggregation cation
 IT Actins
 RL: BIOL (Biological study)
 (aggregation of, divalent cations effect on)
 IT Cations
 (divalent, actin aggregation response to)
 IT Molecular association
 (self-, of actin in divalent cation presence)
 IT 7439-95-4, biological studies 7439-96-5, biological studies
 7440-02-0, biological studies 7440-24-6, biological studies 7440-66-6,
 biological studies 7440-70-2, biological studies
 RL: BIOL (Biological study)
 (actin aggregation response to)
 IT 7439-95-4, biological studies
 RL: BIOL (Biological study)
 (actin aggregation response to)
 RN 7439-95-4 HCPLUS
 CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 17 OF 18 HCPLUS COPYRIGHT 2002 ACS
 AN 1975:39842 HCPLUS
 DN 82:39842
 TI Biochemical and structural studies of actomyosin-like proteins from nonmuscle cells. II. Purification, properties, and membrane association of actin from amebae of Dictyostelium discoideum
 AU Spudich, James A.; Lord, Kathy
 CS Dep. Biochem. Biophys., Univ. California, San Francisco, Calif., USA
 SO J. Biol. Chem. (1974), 249(18), 6013-20
 CODEN: JBCHA3
 DT Journal
 LA English
 CC 6-3 (General Biochemistry)

AB Actomyosin was obtained from the title amebas by the method of M. Clarke and J. A. Spudich (1974), and actin was purified from this prepn. The sp. activity of this actin for activation of heavy meromyosin ATPase was comparable to that of muscle actin. The ameba actin and muscle actin comigrated on Na dodecyl sulfate (I)-acrylamide gels at a rate corresponding to a mol. wt. of apprx. 42,000. The ameba actin formed Mg²⁺ paracrystals with a repeating band pattern of 300-400 .ANG., similar to muscle and platelet actins. Purifn. of ameba membranes by sedimentation equil. on sucrose gradients resulted in an apprx. 3-fold copurifn. of actin. Sepn. of membrane components by I gel electrophoresis established that the myosin and actin components maintained a const. ratio relative to other components in membranes subjected to centrifugation for varying periods of time. Further, MgATP released all of the myosin and apprx. 1/2 of the actin from the membranes. In the absence of MgATP, apprx. 10% of the total cellular actin was recovered with membranes. Thus, apprx. 5% of the actin was assocd. with membranes in a MgATP-stable linkage. This assocn. may be analogous to actin assocn. with z-lines in muscle. A model for nonrandom movement in nonmuscle cells was constructed which is consistent with the above results and with the principles of actin-myosin interaction in sarcomeres.

ST cell membrane Dictyostelium actin; magnesium ATP
Dictyostelium actin; movement Dictyostelium actin

IT Cell membrane
(actin assocd. with, of Dictyostelium discoideum, nonrandom movement model in relation to)

IT Dictyostelium discoideum
(actin of, sepn. and characterization of, nonrandom movement model in relation to)

IT **Actins**
RL: BIOL (Biological study)
(of Dictyostelium discoideum, sepn. and charactization of, nonrandom movement model in relation to)

IT Adenosine 5'-(tetrahydrogen triphosphate), magnesium salt (1:1), magnesium complexes
magnesium, ATP complexes
RL: BIOL (Biological study)
(actin of Dictyostelium discoideum in response to, cell membrane assocn. in relation to)

IT 7439-95-4, biological studies
RL: BIOL (Biological study)
(actin of Dictyostelium discoideum paracrystal formation with)

IT 7439-95-4, biological studies
RL: BIOL (Biological study)
(actin of Dictyostelium discoideum paracrystal formation with)

RN 7439-95-4 HCAPLUS
CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

L31 ANSWER 18 OF 18 HCAPLUS COPYRIGHT 2002 ACS
AN 1971:71735 HCAPLUS
DN 74:71735
TI Polymorphism of F-actin. I. Three forms of paracrystals
AU Kawamura, Masaru; Maruyama Kosca
CS Biol. Inst., Univ. Tokyo, Tokyo, Japan
SO J. Biochem. (Tokyo) (1970), 68(6), 885-99
CODEN: JOBIAO

DT Journal
 LA English
 CC 2 (General Biochemistry)
 AB Polymorphic assemblies of F-actin were studied at acid pH using an electron microscope. Three distinct types of ordered aggregates, designated as TYPE I, II, and III, were found and their basic structural features were described. TYPE I was a net with 2-fold rotational symmetry and the tetragon had rms of 320 .ANG. in length and the angles between the arms were 28.degree. and 152.degree., resp. For the formation of TYPE I, the optimal concn. of KCl and ATP were 0.1-0.2M and 0.4mM at pH 5.0, resp. TYPE II was also a net similar to TYPE I, but the distribution of matter was different from TYPE I, and TYPE II was more rigid in structure. The conditions of formation of TYPE II were not elucidated. TYPE III was a side-by-side aggregate of F-actin similar to that formed in the presence of MgCl₂ (Hanson, 1967), but TYPE III appeared to be somewhat different in shape. ATP or KCl was not necessary for the formation of TYPE III. F-actin showed ATPase [EC 3.6.1.3] activity at acid pH. This ATPase action was discussed in relation to the formation of the TYPE I paracrystal.
 ST polymorphism F actin; actins polymorphism; structure F actin
 IT Crystal structure
 (of actin F)
 IT Phosphatases, adenosine tri-
 (of actin F polymorphic crystals)
 IT Actins
 RL: BIOL (Biological study)
 (polymorphism of F-, structure of)

=> d 130 all hitstr

L30 ANSWER 1 OF 1 HCAPLUS COPYRIGHT 2002 ACS
 AN 1985:109256 HCAPLUS
 DN 102:109256
 TI A new simple method of preparing actin from chicken gizzard
 AU Ebashi, Setsuro
 CS Fac. Med., Univ. Tokyo, Tokyo, 113, Japan
 SO J. Biochem. (Tokyo) (1985), 97(2), 693-5
 CODEN: JOBIAO; ISSN: 0021-924X
 DT Journal
 LA English
 CC 9-6 (Biochemical Methods)
 AB A simple method for prep. actin from chicken gizzard is described. The method involves a series of centrifugations and then acetone treatment, resulting in an acetone powder of the gizzard. The acetone powder is then subjected to further centrifugation steps, and the purity of the resulting actin prepns. is examd. by SDS-polyacrylamide gel electrophoresis and electron microscopic profiles. This method takes advantage of the property of gizzard tropomyosin that it does not form Mg paracrystals readily. The method gives actins with higher specific viscosity than the conventional method, it removes F-actin disaggregating factors, and it gives a yield of usually 15-20 mg of actin.
 ST actin prep chicken gizzard; centrifugation actin prep
 IT Tropomyosins
 RL: PREP (Preparation)
 (actin prep. from chicken gizzard in magnesium presence in relation to)
 IT Chicken
 (actin prep. from gizzard of)
 IT Gizzard
 (actin prep. from, of chicken by centrifugation)
 IT Centrifugation
 (in actin prep. from chicken gizzard)

IT Actins
RL: PREP (Preparation)
(prepn. of, from chicken gizzard by centrifugation)
IT Actins
RL: PREP (Preparation)
(F-, prepn. of, from chicken gizzard by centrifugation)
IT 7439-95-4, biological studies
RL: ANST (Analytical study)
(actin prepn. from chicken gizzard in presence of,
tropomyosin in relation to)
IT 7439-95-4, biological studies
RL: ANST (Analytical study)
(actin prepn. from chicken gizzard in presence of,
tropomyosin in relation to)
RN 7439-95-4 HCPLUS
CN Magnesium (8CI, 9CI) (CA INDEX NAME)

Mg

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L61 ANSWER 1 OF 1 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 1995:166199 BIOSIS
DN PREV199598180499
TI Concentrated Tris Solutions for the Preparation,
Depolymerization and Assay of Actin: Application to Erythroid
Actin.
AU Pinder, Jennifer C.; Sleep, J. A.; Bennett, Pauline M.; Gratzer, W. B.
CS Med. Res. Council Muscle Cell Motility Unit, King's Coll., 26-29 Drury
Lane, London WC2B 5RL UK
SO Analytical Biochemistry, (1995) Vol. 225, No. 2, pp. 291-295.
ISSN: 0003-2697.
DT Article
LA English
AB High concentrations of Tris are effective in dissociating actin
-containing complexes, such as the red cell membrane cytoskeleton. A
preparative procedure for red cell actin is based on the
dissociation of the membrane skeletal complex in a buffer containing 1 M
Tris hydrochloride, followed by gel filtration chromatography in the same
medium. The actin is recovered as the monomer and is fully
native, as judged by its critical concentration of polymerization,
inhibition of DNase I, stimulation of myosin ATPase, and the appearance in
the electron microscope of filaments, both bare and decorated with heavy
meromyosin, and of magnesium ion-induced paracrystals.
The Tris solution causes rapid depolymerization of F-actin with
no denaturation, and the solution of monomeric actin in this
medium is stable for many weeks in the cold; concentrated Tris is more
reliable than guanidinium chloride for the depolymerization of F-
actin in the estimation of total actin concentration by

CC the DNase I inhibition assay.
Biochemical Methods - Proteins, Peptides and Amino Acids *10054
Biochemical Studies - Proteins, Peptides and Amino Acids 10064
Biophysics - Membrane Phenomena *10508
Enzymes - Methods *10804
Blood, Blood-Forming Organs and Body Fluids - Blood Cell Studies *15004
IT Major Concepts
 Blood and Lymphatics (Transport and Circulation); Enzymology
 (Biochemistry and Molecular Biophysics); Membranes (Cell Biology);
 Methods and Techniques
IT Chemicals & Biochemicals
 TRIS; ACTIN; ATPASE; DNASE I
IT Miscellaneous Descriptors
 DNASE I INHIBITION ASSAY; MYOSIN ATPASE; RED CELL MEMBRANE CYTOSKELETON
RN 77-86-1Q (TRIS)
126-72-7Q (TRIS)
17096-07-0Q (TRIS)
132579-20-5 (ACTIN)
9000-83-3 (ATPASE)
9003-98-9 (DNASE I)

=> fil medline
FILE 'MEDLINE' ENTERED AT 17:18:51 ON 11 FEB 2002

FILE LAST UPDATED: 9 FEB 2002 (20020209/UP). FILE COVERS 1958 TO DATE.

On April 22, 2001, MEDLINE was reloaded. See HELP RLOAD for details.

MEDLINE now contains IN-PROCESS records. See HELP CONTENT for details.

MEDLINE is now updated 4 times per week. A new current-awareness alert frequency (EVERYUPDATE) is available. See HELP UPDATE for more information.

MEDLINE thesauri in the /CN, /CT, and /MN fields incorporate the MeSH 2001 vocabulary. Enter HELP THESAURUS for details.

The OLDMEDLINE file segment now contains data from 1958 through 1965. Enter HELP CONTENT for details.

Left, right, and simultaneous left and right truncation are available in the Basic Index. See HELP SFIELDS for details.

THIS FILE CONTAINS CAS REGISTRY NUMBERS FOR EASY AND ACCURATE SUBSTANCE IDENTIFICATION.

=> d all tot

L84 ANSWER 1 OF 4 MEDLINE
AN 92256699 MEDLINE
DN 92256699 PubMed ID: 1581508
TI Linear dichroism of acrylodan-labeled tropomyosin and myosin subfragment 1 bound to actin in myofibrils.
AU Szczesna D; Lehrer S S
CS Department of Muscle Research, Boston Biomedical Research Institute, Massachusetts 02114.
NC HL-22461 (NHLBI)
SO BIOPHYSICAL JOURNAL, (1992 Apr) 61 (4) 993-1000.
Journal code: A5S; 0370626. ISSN: 0006-3495.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199206
ED Entered STN: 19920626
Last Updated on STN: 19920626

AB Entered Medline: 19920618

AB Muscle contraction can be activated by the binding of myosin heads to the thin filament, which appears to result in thin filament structural changes. In vitro studies of reconstituted muscle thin filaments have shown changes in tropomyosin-actin geometry associated with the binding of myosin subfragment 1 to actin. Further information about these structural changes was obtained with fluorescence-detected linear dichroism of tropomyosin, which was labeled at Cys 190 with acrylodan and incorporated into oriented ghost myofibrils. The fluorescence from three sarcomeres of the fibril was collected with the high numerical aperture objective of a microscope and the dichroic ratio, R (0/90 degrees), for excitation parallel/perpendicular to the fibril, was obtained, which gave the average probe dipole polar angle, Theta. For both acrylodan-labeled tropomyosin bound to actin in fibrils and in Mg²⁺ paracrystals, Theta congruent to 52 degrees +/- 1.0 degrees, allowing for a small degree of orientational disorder. Binding of myosin subfragment 1 to actin in fibrils did not change Theta; i.e., the orientation of the rigidly bound probe on tropomyosin did not change relative to the actin axis. These data indicate that myosin subfragment 1 binding to actin does not appreciably perturb the structure of tropomyosin near the probe and suggest that the geometry changes are such as to maintain the parallel orientation of the tropomyosin and actin axes, a finding consistent with models of muscle regulation. Data are also presented for effects of MgADP on the orientation of labeled myosin subfragment 1 bound to actin in myofibrils.

CT Check Tags: Animal; Support, Non-U.S. Gov't; Support, U.S. Gov't, Non-P.H.S.; Support, U.S. Gov't, P.H.S.

2-Naphthylamine: AA, analogs & derivatives

Actins: CH, chemistry

Adenosine Diphosphate

Binding Sites

Biophysics

Fluorescent Dyes

Models, Chemical

Myofibrils: CH, chemistry

*Myosin Subfragments: CH, chemistry

Rabbits

Spectrometry, Fluorescence

*Tropomyosin: CH, chemistry

RN 58-64-0 (Adenosine Diphosphate); 86636-92-2 (acrylodan); 91-59-8 (2-Naphthylamine)

CN 0 (Actins); 0 (Fluorescent Dyes); 0 (Myosin Subfragments); 0 (Tropomyosin)

L84 ANSWER 2 OF 4 MEDLINE

AN 92223214 MEDLINE

DN 92223214 PubMed ID: 1839660

TI [Purification and biochemical characteristics of actin from the rat malignancy sarcoma-45].

Ochistka i biokhimicheskaya kharakteristika aktina iz zлокачественнои опухоли крыс саркома-45.

AU Senchuk V V; Pikulev A T; Dashkevich I N

SO BIORHIMIIA, (1991 Dec) 56 (12) 2235-43.

Journal code: A28; 0372667. ISSN: 0320-9725.

CY USSR

DT Journal; Article; (JOURNAL ARTICLE)

LA Russian

FS Priority Journals

EM 199205

ED Entered STN: 19920607

Last Updated on STN: 19920607

Entered Medline: 19920521

AB Actin was purified from rat sarcoma-45 by using affinity chromatography on DNase I agarose. Actin was detected in the soluble and cytoskeletal fractions. The molecular mass of the

protein was found to be equal to 45 kDa. The tumour actin specifically reacted with the antibody against skeletal muscle actin, inhibited the DNAase I activity and activated in the fibrillar state Mg(2+)-ATPases of sarcoma-45 and skeletal muscle myosins. The activating effect of the tumour protein was lower than that of its skeletal muscle counterpart. V8-protease peptide mapping revealed a similarity between tumour and brain actins. Sarcoma-45 actin was found to contain beta- and gamma-actin isoforms and an unusual isoform which appeared to be more acidic than the alpha-actin isoform.

CT Check Tags: Animal

Actins: IP, isolation & purification

*Actins: ME, metabolism

Ca(2+) Mg(2+)-ATPase: ME, metabolism

Deoxyribonuclease I: ME, metabolism

Electrophoresis, Gel, Two-Dimensional

Electrophoresis, Polyacrylamide Gel

Myosin: ME, metabolism

Pancreas: EN, enzymology

Rats

*Sarcoma, Experimental: ME, metabolism

CN 0 (Actins); 0 (Myosin); EC 3.1.21.1 (Deoxyribonuclease I); EC 3.6.1.- (Ca(2+) Mg(2+)-ATPase)

L84 ANSWER 3 OF 4 MEDLINE

AN 84000622 MEDLINE

DN 84000622 PubMed ID: 6137243

TI Comparison of the properties of two kinds of preparations of human blood platelet actin with sarcomeric actin.

AU Coue M; Landon F; Olomucki A

SO BIOCHIMIE, (1982 Mar) 64 (3) 219-26.

Journal code: A14; 1264604. ISSN: 0300-9084.

CY France

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 198311

ED Entered STN: 19900319

Last Updated on STN: 19900319

Entered Medline: 19831123

AB A new procedure of purification of actin from human blood platelets was used. This method starting from acetone powder of whole platelets gives a much higher yield than the one previously described (actin I) (Landon et al. (1977) Eur. J. Biochem., 81, 571-577). This actin II preparation has the same reduced viscosity as skeletal muscle actin, while the reduced viscosity of actin I preparation is about 1/10 of this value. Moreover actin I has the form of very short filaments as shown by electron microscopy. After an extra step of purification actin I, when polymerized, acquired a high reduced viscosity. We confirmed that platelet and sarcomeric actins are similar in their polymerization properties and their ability to activate muscular myosin. A circular dichroism study showed that the overall conformation of both actins are similar, but the environment of their aromatic chromophores is different.

CT Check Tags: Animal; Comparative Study; Human; Support, Non-U.S. Gov't

Actins: BL, blood

Actins: IP, isolation & purification

*Actins: PD, pharmacology

Adenosinetriphosphatase: ME, metabolism

*Blood Platelets: AN, analysis

Ca(2+) Mg(2+)-ATPase

Circular Dichroism

Enzyme Activation

Macromolecular Systems

*Myofibrils: AN, analysis

Protein Conformation
 Rabbits
 *Sarcomeres: AN, analysis
 Viscosity
 CN 0 (Actins); 0 (Macromolecular Systems); EC 3.6.1.- (Ca(2+)
 Mg(2+)-ATPase); EC 3.6.1.3 (Adenosinetriphosphatase)

L84 ANSWER 4 OF 4 MEDLINE
 AN 78084385 MEDLINE
 DN 78084385 PubMed ID: 145944
 TI Human platelet actin. Evidence of beta and gamma forms and similarity of properties with sarcomeric actin.
 AU Landon F; Huc C; Thome F; Oriol C; Olomucki A
 SO EUROPEAN JOURNAL OF BIOCHEMISTRY, (1977 Dec) 81 (3) 571-7.
 Journal code: EMZ; 0107600. ISSN: 0014-2956.
 CY GERMANY, WEST: Germany, Federal Republic of
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Priority Journals
 EM 197803
 ED Entered STN: 19900314
 Last Updated on STN: 19900314
 Entered Medline: 19780321
 AB Human blood platelet actin was purified using 30% sucrose to extract actomyosin and potassium iodide to dissociate actomyosin and to depolymerize actin. Pure actin thus obtained resembles skeletal muscle actin in its polymerization properties, CD spectra and ability to activate myosin myosin Mg²⁺-ATPase. Isoelectric focusing gel analysis shows that human blood platelet actin exists in beta and gamma forms. The ratio of beta to gamma forms is of 5 in purified actin, in whole cell extract and in all the fractions studied.
 CT Check Tags: Animal; Comparative Study; Human
 *Actins
 Actins: BL, blood
 Actins: IP, isolation & purification
 Adenosinetriphosphatase: ME, metabolism
 *Blood Platelets: AN, analysis
 Macromolecular Systems
 Molecular Weight
 Muscles
 Myosin
 Organ Specificity
 Protein Conformation
 Rabbits

CN 0 (Actins); 0 (Macromolecular Systems); 0 (Myosin); EC 3.6.1.3
 (Adenosinetriphosphatase)

=> d his

(FILE 'HOME' ENTERED AT 16:22:44 ON 11 FEB 2002)
 SET COST OFF

FILE 'REGISTRY' ENTERED AT 16:22:56 ON 11 FEB 2002
 E MAGNESIUM/CN

L1 1 S E3
 E MAGNESIUM CHLORIDE/CN
 L2 1 S E3

FILE 'HCAPLUS' ENTERED AT 16:23:32 ON 11 FEB 2002
 E ACTIN/CT
 E E3+ALL

L3 1 S E1
 E E2+ALL
 L4 19413 S E2

L5 1133 S E2 (L) G
 L6 1134 S L3,L5
 L7 4805 S ACTIN (L) G
 L8 4805 S L6,L7
 L9 36371 S ACTIN
 L10 168657 S L1 OR L2
 L11 47463 S MAGNESIUM CHLORIDE OR MGCL2 OR MAGNESIU
 L12 895 S L3-L9 AND L10,L11
 L13 36 S L12 AND ?CRYSTAL?
 L14 130 S L3-L9 AND (PARACRYSTAL? OR PARA(L)?CRYSTAL?)
 L15 24 S L14 AND L10,L11
 L16 36 S L13,L15
 L17 125 S ACTINS/CW (L) PREP/RL
 L18 1 S L16 AND L17
 E HARTMAN J/AU
 L19 22 S E3,E11
 E HARTMAN JAMES/AU
 L20 9 S E3,E8,E9
 E MALIK F/AU
 L21 193 S E3-E12
 E SAKOZIC R/AU
 E SAKOWIC R/AU
 L22 23 S E10,E12
 E FINER J/AU
 L23 16 S E3,E6,E9,E10
 L24 7 S L3-L9,L17 AND L19-L23
 L25 13 S L16 AND (FORMATION OR ISOLATION OR CHARACTERIZATION OR POLYMER)
 L26 12 S L25 NOT ASCARIS/TI
 L27 13 S L18,L26
 L28 12 S L27 AND (MAGNESIUM OR MGCL2 OR MG###)
 L29 19 S L24,L28 AND L1-L28
 L30 1 S L29 AND L17
 L31 18 S L29 NOT L30

FILE 'REGISTRY' ENTERED AT 16:44:38 ON 11 FEB 2002

FILE 'HCAPLUS' ENTERED AT 16:44:50 ON 11 FEB 2002
 L32 36371 S L3-L9,L17
 L33 31 S L32 AND COMBINATOR?
 L34 22 S L32 AND HIGH(L)(THROUGHPUT OR THROUGH PUT)
 E .BETA.-ACTIN/CT
 E E6+ALL
 L35 732 S L32 AND ?CRYSTAL?
 L36 3 S L35 AND L33,L34
 L37 3 S L36 NOT L30,L31
 L38 1 S L35 AND SOLID(L) PHASE
 E COMBINATORIAL/CT
 L39 6641 S E5+NT OR E6+NT
 E E5+ALL
 L40 125 S E6+NT
 E E8+ALL
 L41 28189 S E2+NT
 L42 20 S L32 AND L39-L41
 L43 20 S L42 NOT L30,L31
 L44 2338 S REACTION+NT/CT AND L32
 L45 32 S L35 AND L44
 L46 75 S L10,L11 AND L44
 L47 292 S (MAGNESIUM OR MGCL2 OR MG###) AND L44
 L48 5 S L45 AND L46,L47
 L49 3 S L48 NOT L30,L31

FILE 'BIOSIS' ENTERED AT 16:55:02 ON 11 FEB 2002

L50 49275 S ACTIN
 L51 579 S L50 AND L1,L2
 L52 80 S L50 AND (MAGNESIUM OR MG)()CHLORIDE
 L53 237 S L50 AND MGCL2

L54 2524 S L50 AND MG##
 L55 2744 S L51-L54
 L56 70 S L55 AND ?CRYSTALS
 L57 1 S L56 AND SUSPENSION/TI
 L58 1405 S L50 AND MAGNESIUM
 L59 40 S L58 AND ?CRYSTALS
 L60 8 S L59 NOT L56
 L61 1 S L60 AND PREPARATION
 L62 7 S L50 AND (HARTMAN J? OR MALIK F? OR SAKOWICZ R? OR FINER J?) /A

FILE 'BIOSIS' ENTERED AT 17:02:27 ON 11 FEB 2002

FILE 'MEDLINE' ENTERED AT 17:02:46 ON 11 FEB 2002
 L63 38076 S L9
 E ACTIN/CT
 E E3+ALL
 E E2+ALL
 L64 22430 S E11/CT,CN
 L65 38076 S L63,L64
 L66 774 S L65 AND L1,L2
 L67 2980 S L65 AND ((MAGNESIUM OR MG) ()CHLORIDE OR MGCL2 OR MG### OR MAG
 L68 82 S L66,L67 AND ?CRYSTALS
 L69 1 S L68 AND DEPOLYMERIZATION/TI AND DIVALENT METAL CHLORIDE/TI
 L70 652 S (ACTINS(L)IP)/CT
 L71 112 S L70 AND L66,L67
 L72 88 S L64/MAJ AND L71
 L73 7 S L72 NOT AB/FA
 L74 81 S L72 NOT L73
 SEL DN 24 72
 L75 2 S E1-E4 AND L74
 L76 3 S L69,L75
 L77 3 S L63-L75 AND L76
 L78 1579 S L65 AND SARCOM?
 L79 72 S L78 AND L66,L67
 L80 1 S L79 AND L68
 L81 3 S L70 AND L79
 L82 4 S L80,L81
 L83 3 S L82 NOT SARCOMA
 L84 4 S L81,L83

FILE 'MEDLINE' ENTERED AT 17:18:51 ON 11 FEB 2002

FILE 'WPIX' ENTERED AT 17:18:58 ON 11 FEB 2002
 L85 497 S ACTIN
 E MAGNESIUM CHLORIDE/DCN
 E E3+ALL
 L86 6218 S E2 OR 1801/DRN
 L87 236682 S MGCL2 OR (MG OR MAGNESIUM) ()CHLORIDE OR MG### OR MAGNESIUM OR
 L88 34 S L85 AND L86,L87
 L89 0 S L88 AND ?CRYSTALS
 L90 1 S L88 AND SARCOM?
 E SARCOM
 L91 13 S E20-E26
 L92 4 S L91 AND L85
 L93 0 S (PARACRYSTALS? OR PARA CRYSTALS?) AND L85
 L94 ,4 S ?CRYSTALS? AND L85